

ab65311

Cytochrome c Releasing Apoptosis Assay Kit

Instructions for Use

For the rapid, sensitive and accurate detection of Cytochrome c translocation from Mitochondria into Cytosol during Apoptosis in cells and tissues

This product is for research use only and is not intended for diagnostic use.

PLEASE NOTE: With the acquisition of BioVision by Abcam, we have made some changes to component names and packaging to better align with our global standards as we work towards environmental-friendly and efficient growth. You are receiving the same high-quality products as always, with no changes to specifications or protocols.

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1. Overview

Cytochrome c plays an important role in apoptosis. The protein is located in the space between the inner and outer mitochondrial membranes. An apoptotic stimulus triggers the release of cytochrome c from the mitochondria into cytosol where it binds to Apaf-1. The cytochrome c/Apaf-1 complex activates caspase-9, which then activates caspase-3 and other downstream caspases.

Abcam's Cytochrome c Releasing Apoptosis Assay Kit provides an effective means for detecting cytochrome c translocation from mitochondria into cytosol during apoptosis. The kit provides unique formulations of reagents to isolate a highly enriched mitochondria fraction from cytosol. The procedure is so simple and easy to perform; no ultracentrifugation is required and no toxic chemicals are involved. Cytochrome c releasing from mitochondria into cytosol is then determined by Western blotting using the cytochrome c antibody provided in the kit.

2. Protocol Summary

3. Components and Storage

A. Kit Components

Item	Quantity
Mitochondria Extraction Buffer I/Mitochondria Extraction Buffer	10 mL
5X Cytosol Extraction Buffer I/5X Cytosol Extraction Buffer	20 mL
DTT II/DTT (1 M)	100 μL
Protease Inhibitor Cocktail I/500X Protease Inhibitor Cocktail (Lyophilized)	1 vial
Anti-Mouse Cyt C Antibody/Anti- Cytochrome c Mouse mAb	100 μL

^{*} Store kit at -20°C.

- Be sure to keep all buffers on ice at all times during the experiment.
- Read the entire protocol before beginning the procedure.
- After opening the kit, store buffers at +4°C. Store the Anti-Mouse Cyt C Antibody/antibody, Protease Inhibitor Cocktail I/Protease Inhibitor Cocktail, and DTT II/DTT at -20°C.

Protease Inhibitor Cocktail I/PROTEASE INHIBITOR COCKTAIL: Add 250 µL DMSO before use.

Mitochondria Extraction Buffer I/MITOCHONDRIA EXTRACTION BUFFER MIX: Before use, prepare just enough Mitochondria Extraction Buffer I/Mitochondria Extraction Buffer Mix for your experiment: Add 2 μ L Protease Inhibitor Cocktail I/Protease Inhibitor cocktail and 1 μ L DTT II/DTT to 1 mL of Mitochondria Extraction Buffer I/Mitochondria Extraction Buffer.

Cytosol Extraction Buffer I/CYTOSOL EXTRACTION BUFFER MIX: Dilute the 5X Cytosol Extraction Buffer I/5X Cytosol Extraction Buffer to 1X buffer with ddH $_2$ O. Before use, prepare just enough Cytosol Extraction Buffer I/Cytosol Extraction Buffer Mix for your experiment: Add 2 μ L Protease Inhibitor Cocktail I/Protease Inhibitor cocktail and 1 μ L DTT II/DTT to 1 mL of 1X Cytosol Extraction Buffer I/Cytosol Extraction Buffer.

B. Additional Materials Required

- Microcentrifuge
- Pipettes and pipette tips

- Dounce tissue grinder
- Orbital shaker

4. Assay Protocol

- **1.** Induce apoptosis in cells by desired method. Concurrently incubate a control culture *without* induction.
- **2.** Collect cells (5 x 10^7) by centrifugation at 600 x g for 5 minutes at 4° C.
- Wash cells with 10 mL of ice-cold PBS. Centrifuge at 600 x g for 5 minutes at 4°C. Remove supernatant.
- 4. a) Cells: Re-suspend cells with 1ml of 1X Cytosol Extraction Buffer I/Cytosol Extraction Buffer Mix containing DTT II/DTT and Protease Inhibitor Cocktail I/Protease Inhibitors. Incubate on ice for 10 minutes.
 - b) Frozen tissue may be suitable (although not tested) but would recommend fresh. If you must use frozen: washing the tissue with ice cold PBS and then resuspend each 10 mg of tissue in 1ml of Cytosol Extraction Buffer I/cytosol extraction buffer.
- 5. Homogenize cells in an ice-cold Dounce tissue grinder. Perform the task with the grinder on ice. We recommend 30-50 passes with the grinder; however, efficient homogenization may depend on the cell type.

Notes:

a) To check the efficiency of homogenization, pipette 2-3 μ L of the homogenized suspension onto a coverslip and observe

under a microscope. A shiny ring around the nuclei indicates that cells are still intact. If 70-80% of the nuclei do not have the shiny ring, proceed to step 7. Otherwise, perform 10-20 additional passes using the Dounce tissue grinder.

- b) Excessive homogenization should also be avoided, as it can cause damage to the mitochondrial membrane which triggers release of mitochondrial components.
- **6.** Transfer homogenate to a 1.5 mL microcentrifuge tube, and centrifuge at 700 x g for 10 minutes at 4°C.
- 7. Collect supernatant into a fresh 1.5 mL tube, and centrifuge at 10,000 x g for 30 minutes at 4°C. Collect supernatant as Cytosolic Fraction.
- 8. Re-suspend the pellet in 100 μL Mitochondria Extraction Buffer I/Mitochondrial Extraction Buffer Mix containing DTT II/DTT and Protease Inhibitor Cocktail I/protease inhibitors (as prepared in section A), vortex for 10 seconds and save as Mitochondrial Fraction.
- 9. Load 10 µg each of the cytosolic and mitochondrial fractions isolated from un-induced and induced cells on a 12% SDS-PAGE. Then proceed with standard Western blot procedure and probe with Anti-Mouse Cyt C Antibody/cytochrome c antibody (1:200 dilution is recommended).

Note:

The Anti-Mouse Cyt C Antibody/anti-Cytochrome c antibody is a mouse monoclonal antibody that reacts with denatured human, mouse, and rat cytochrome c.

For further technical questions please do not hesitate to contact us by email (technical@abcam.com) or phone (select "contact us" on www.abcam.com for the phone number for your region).



Technical Support

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